HUMAN IL8 KIT

PROTOCOL

Part # 62HIL08PET

Test size: 1 x 96 tests (62HIL08PET) - assay volume: 20 µL

Revision: 07 (May 2020)

Store at: ≤-60°C

This product is intended for research purposes only. The product is not intended to be used for therapeutic or diagnostic purposes.

ASSAY PRINCIPLE

Cisbio Bioassays' human IL8 assay is only intended for the quantitative measurement of IL8 in supernatant using HTRF® technology. The assay is compatible with human samples, and is highly specific for IL8.

IL8 is detected in a sandwich assay format using 2 different specific antibodies, one labeled with Europium Cryptate (donor) and the second with d2 (acceptor).

The detection principle is based on HTRF® technology. When the labelled antibodies bind to the same antigen, the excitation of the donor with a light source (laser or flash lamp) triggers a Fluorescence Resonance Energy Transfer (FRET) towards the acceptor, which in turn fluoresces at a specific wavelength (665 nm). The two antibodies bind to the IL8 present in the sample, thereby generating FRET. Signal intensity is proportional to the number of antigen-antibody complexes formed and therefore to the IL8 concentration. (Fig. 1).

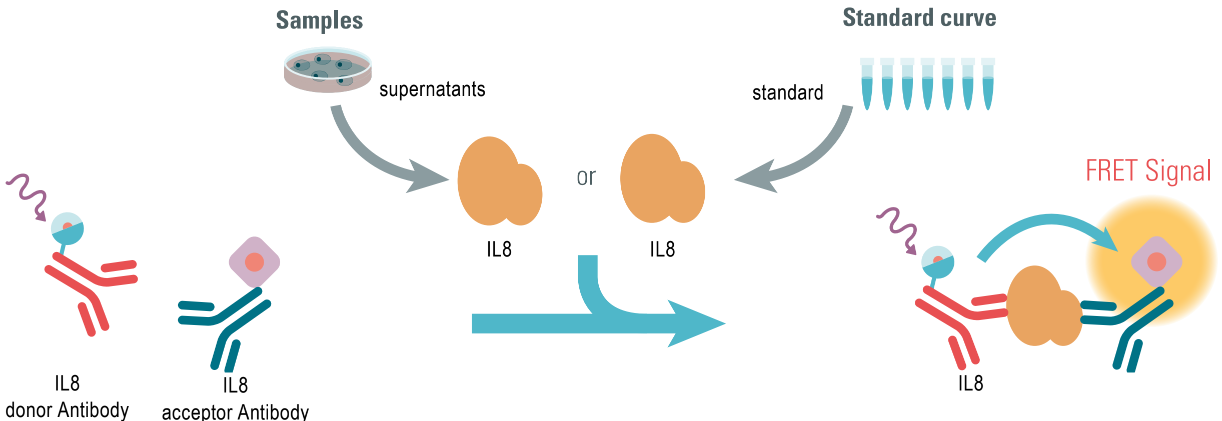


Figure 1: Principle of HTRF IL8 sandwich assay.

PROTOCOL AT A GLANCE

ADD

16 µL
Standard
or Sample

+

4 µL
of pre-mixed IL8 Antibodies

Incubate
1 hours*
at room
temperature

READ

*Following incubation, the signal remains stable over a period of 24 hours.

ANALYSE

HTRF Signal

[Cytokine] pg/mL

White 384-well low volume plate

Make sure to use the set-up for Eu^{3+} Cryptate.
For more information about set-up and compatible HTRF® readers, please visit our website at www.cisbio.com/readers

MATERIALS:

KIT COMPONENTS	1 X 96 TESTS - CAT # 62HIL08PET
IL8 Standard Lyophilized	1 vial
IL8 Eu Cryptate Antibody Frozen	1 vial - 10 µL
IL8-d2 Antibody Frozen	1 vial - 10 µL
Diluent* #5 5X	1 vial 2 mL
Detection Buffer** #3 ready-to-use	1 vial - 0.5 mL
Plate	1 plate HTRF 96-well low volume plate

* To prepare working standard solutions, culture medium can be an alternative the diluent.
** The Detection Buffer is used to prepare working solutions of acceptor and donor reagents.

FOR READING, AN HTRF®-CERTIFIED READER IS NEEDED.

For a list of HTRF-compatible readers and set-up recommendations, please visit www.cisbio.com/readers

PURCHASE SEPARATELY

96-well or 384-well small volume (SV) detection microplates - For more information about microplate recommendations, please visit our website at: www.cisbio.com/content/microplates-recommendations

STORAGE AND STABILITY

Store the kit at ≤-60°C. Under proper storage conditions, reagents are stable until the expiry date indicated on the label.

Reagents

Once reconstituted, standard stock solution may be frozen, and can be thawed only once..
Once thawed, antibody solutions can be frozen once.
To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at ≤-60°C.

Volume of standard and antibody aliquots should not be under 10 µL.

Thawed diluent and detection buffer can be stored at 2-8°C on your premises.

REAGENT PREPARATION

BEFORE YOU BEGIN:

- It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature.
- Before use, allow all kit's reagents to warm up at room temperature then
  - homogeneize buffer and diluent with a vortex
  - centrifuge (NEVER vortex) the antibodies to gather all liquid at the bottom of the vial
- It is recommended to filter buffers before use.
- Antibody solutions must be prepared in individual vials and can be mixed prior to dispensing.

TAKE CARE TO PREPARE STOCK AND WORKING SOLUTIONS ACCORDING TO THE DIRECTIONS FOR THE KIT SIZE YOU HAVE PURCHASED.

TO PREPARE DILUENT, STANDARD & ANTIBODY STOCK SOLUTIONS:

1 X 96 TESTS	
IL8 Eu Cryptate antibody	
Thaw the IL8 Eu Cryptate antibody. Centrifuge. This stock solution can be frozen and stored at ≤-60°C.	
IL8 d2 antibody	
Thaw the IL8 d2 antibody. Centrifuge. This stock solution can be frozen and stored at ≤-60°C.	
IL8 Standard	
Reconstitute the IL8 standard with distilled water. Volume of reconstitution is indicated on the vial label. The reconstituted standard solution can be frozen and stored at -60°C or below.	
Diluent	
Dilute 5-fold the 5 X diluent #5 with distilled water: homogenize the 5 X diluent #5 with a vortex and add 1 volume of stock solution in 4 volumes of distilled water e.g. 1 mL of diluent + 4 mL of distilled water Mix gently after dilution.	4 vol. 1 vol. 5X

TO PREPARE WORKING ANTIBODY SOLUTIONS:

Each well requires 4 µL of pre-mixed IL8 antibodies. Prepare the two antibody solutions in separate vials.

1 X 96 TESTS	
IL8 Eu Cryptate antibody	
Dilute the stock solution (thawed reagent) of IL8 Eu Cryptate-antibody with detection buffer #3: Add 200µL of detection buffer directly in the thawed Eu Cryptate-antibody stock solution.	200 µL
IL8-d2 antibody	
Dilute the stock solution (thawed reagent) of IL8 d2 antibody with detection buffer #3: Add 200µL of detection buffer directly in thawed d2 antibody stock solution.	200 µL
Antibody mix	
Pre-mix the two ready-to-use antibody solutions just prior to dispensing the reagents: e.g. 210 µL of d2 antibody + 210 µL of Eu Cryptate antibody.	1 vol. 1 vol.

TO PREPARE WORKING STANDARD SOLUTIONS:

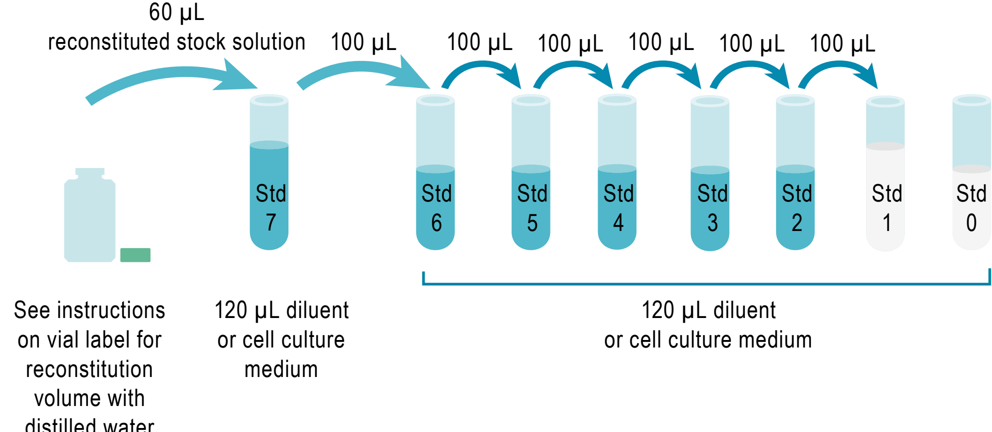
- Each well requires 16 µL of standard.
- Serially dilute the standard stock solution with with diluent #5. or with the cell culture medium used to prepare your samples supplemented with BSA or 10% FCS.
- Due to the stability of the IL8, it is mandatory to prepare the standard curve just before the assay.
- In order to check for a potential interference effect from your own assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent.
- In order to counteract any standard sticking we recommend changing tips between each dilution.

A recommended standard dilution procedure is listed and illustrated below:

1. Reconstitute the standard vial with the volume indicated on the vial label using distilled water.

2. Prepare the following dilutions:

• Dilute the reconstituted standard stock solution 3-fold with diluent or with cell culture medium.
In practice: take 60 µL of stock solution and add it to 120 µL of diluent or cell culture medium. Mix gently. This yields the high standard (Std 7: 4000 pg/mL) for the top of the curve.
•.Use the high standard (Std 7) to prepare the standard curve using serial dilutions as follows:
- Dispense 120 µL of diluent or cell culture medium into each vial from Std 6 to Std 0
- Add 100 µL of standard to 120 µL of diluent or cell culture medium, mix gently and repeat the serial dilution to make standard solutions: std6, std5, std4, std3, std2, std1
This will create 7 standards for the analyte. Std 0 (Negative control) is diluent or appropriate culture medium alone.

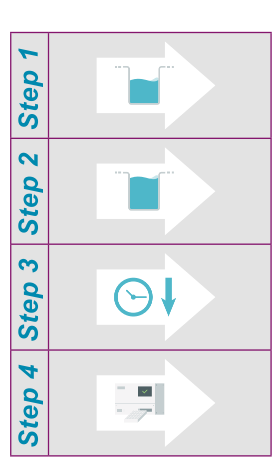


STANDARD	SERIAL DILUTIONS	WORKING SOLUTIONS
Standard Stock solution	Reconstitute the vial following the indications given on the vial label	12 ng/mL
Standard 7	60 µL reconstituted standard stock solution + 120 µL diluent	4000 pg/mL
Standard 6	100 µL Standard 7 + 120 µL diluent	1818.2 pg/mL
Standard 5	100 µL Standard 6 + 120 µL diluent	826.4 pg/mL
Standard 4	100 µL Standard 5 + 120 µL diluent	375.7 pg/mL
Standard 3	100 µL Standard 4 + 120 µL diluent	170.8 pg/mL
Standard 2	100 µL Standard 3 + 120 µL diluent	77.6 pg/mL
Standard 1	100 µL Standard 2 + 120 µL diluent	35.3 pg/mL
Standard 0	120 µL diluent	0

TO PREPARE SAMPLES:

- Each well requires 16 µL of sample.
- Just after their collection, put the samples at 4°C and test them immediately. For later use, samples should be dispensed into disposable plastic vials and stored at ≤-60°C. Avoid multiple freeze/thaw cycles.
- All samples with a concentration above the highest standard (Std 7) must be diluted in diluent #5 or in your cell culture medium.

ASSAY PROTOCOL



	STANDARD (STD 0 - STD 7)	SAMPLES
Step 1	Dispense 16 µL of each IL8 standard (Std 0 - Std 7) into each standard well.	Dispense 16 µL of each sample into each sample well.
Step 2	Dispense 4 µL of pre-mixed IL8 antibodies working solution into all wells.	
Step 3	Seal the plate and incubate at room temperature. *Following incubation, the signal remains stable over a period of 24 hours.	
Step 4	Remove the plate sealer and read on an HTRF® compatible reader.	

	1	2	3	4	5	6
A	16 µL Std 0 4 µL pre-mixed IL8 antibodies	Repeat Well A1	Repeat Well A1	16 µL Sample 1 4 µL pre-mixed IL8 antibodies	Repeat Well A4	Repeat Well A4
B	16 µL Std 1 4 µL pre-mixed IL8 antibodies	Repeat Well B1	Repeat Well B1	16 µL Sample 2 4 µL pre-mixed IL8 antibodies	Repeat Well B4	Repeat Well B4
C	16 µL Std 2 4 µL pre-mixed IL8 antibodies	Repeat Well C1	Repeat Well C1	16 µL Sample 3 4 µL pre-mixed IL8 antibodies	Repeat Well C4	Repeat Well C4
D	16 µL Std 3 4 µL pre-mixed IL8 antibodies	Repeat Well D1	Repeat Well D1	16 µL Sample ... 4 µL pre-mixed IL8 antibodies	Repeat Well D4	Repeat Well D4
E	16 µL Std 4 4 µL pre-mixed IL8 antibodies	Repeat Well E1	Repeat Well E1	16 µL Sample ... 4 µL pre-mixed IL8 antibodies	Repeat Well E4	Repeat Well E4
F	16 µL Std 5 4 µL pre-mixed IL8 antibodies	Repeat Well F1	Repeat Well F1	16 µL Sample ... 4 µL pre-mixed IL8 antibodies	Repeat Well F4	Repeat Well F4
G	16 µL Std 6 4 µL pre-mixed IL8 antibodies	Repeat Well G1	Repeat Well G1	16 µL Sample ... 4 µL pre-mixed IL8 antibodies	Repeat Well G4	Repeat Well G4
H	16 µL Std 7 4 µL pre-mixed IL8 antibodies	Repeat Well H1	Repeat Well H1	16 µL Sample ... 4 µL pre-mixed IL8 antibodies	Repeat Well H4	Repeat Well H4

DATA REDUCTION & INTERPRETATION

1. Calculate the ratio of the acceptor and donor emission signals for each individual well.

$$\text{Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10^4$$

2. **Calculate the delta ratio of the acceptor and donor emission signals for each individual well.** The Standard 0 (Negative control) plays the role of an internal assay control.

delta Ratio = Ratio Standard or sample - Ratio Standard 0

3. **Calculate the % CVs.** The mean and standard deviation can then be worked out from ratio replicates.

$$\text{CV (\%)} = \frac{\text{Standard deviation}}{\text{Mean Ratio}} \times 100$$

For more information about data reduction, please visit www.cisbio.com/data-reduction

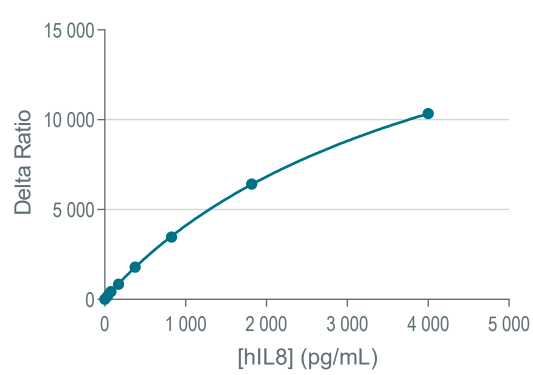
RESULTS

This data must not be substituted for the data obtained in the laboratory and should be considered only as an example.Results may vary from one HTRF® compatible reader to another.

Standard curve fitting with the 4 Parameter Logistic (4PL $1/y^2$)* model

* For more information about curve fitting please visit www.cisbio.com/4PL-regression

		Ratio (1)	delta R (2)	CV% (3)
Standard 0	Negative control	648	0	3%
Standard 1	35.3 pg/mL	835	186	3%
Standard 2	77.6 pg/mL	1079	431	7%
Standard 3	170.8 pg/mL	1493	845	2%
Standard 4	375.7 pg/mL	2446	1789	2%
Standard 5	826.4 pg/mL	4119	3471	1%
Standard 6	1818.2 pg/mL	7072	6423	1%
Standard 7	4000 pg/mL	10994	10346	1%



ANALYTICAL ASSAY PERFORMANCE

	Diluent	DMEM	RPMI
Assay range (pg/mL**)	32 pg/mL to 4000 pg/mL		
Limit of detection (LoD*) = Std 0 mean + 2 SD	6.1 pg/mL	10.9 pg/mL	8.5 pg/mL
Limit of quantification (LoQ*)	32 pg/mL		
Incubation time	at room temperature		

**NIBSC (89/520) value (IU/mL) = 0,01 x HTRF hIL8 value (pg/mL)

*the analytical sensitivity was calculated from data obtained with an HTRF compatible reader after 1 hours of incubation, this may vary from one HTRF compatible reader to another.

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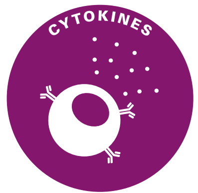


human IL8 kits

SHORT PROTOCOL

Part # 62HIL08PET

Test size: 1 x 96 tests (62HIL08PET) - assay volume: 20 µL

1

Allow all reagents to warm up to room temperature for at least 30 minutes before the assay.

2

Prepare the d2 and Eu cryptate antibody working solutions.
Centrifuge the stock solutions before dilution.
Dilute the stock solution (thawed reagent) of d2 and Eu cryptate antibodies with detection buffer:
Add 200 µL of detection buffer directly in thawed d2 and Eu cryptate antibodies vials.

3

Prepare the mixed antibody solution.
Add 1 volume of 1X Eu cryptate antibody working solution to 1 volume of 1X d2 antibody working solution.

4

Prepare the 1 X diluent working solution.
Dilute 5-fold the 5 X diluent stock solution with distilled water.
(e.g. add 1 mL of 5X diluent to 4 mL of distilled water).

5

Prepare the Standard max (Std 7).
First, reconstitute the lyophilized standard with distilled water. The volume of reconstitution is indicated on the vial label.
Then dilute 3-fold this standard stock solution (e.g. take 60 µL of standard stock solution and add it to 120 µL of diluent or culture medium).

6

Prepare the standard curve.
Make serial dilutions from Std 7 to Std 1 in diluent or in your culture medium (e.g. take 100 µL of Std 7 and add it to 120 µL of diluent or culture medium, this will create Std 6. Repeat this process from Std 6 to Std 1).

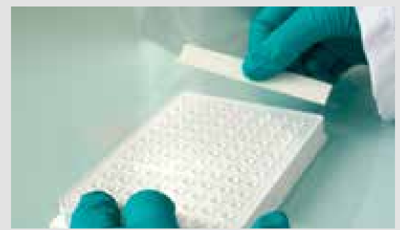
7

Dispense standard curve and samples.
Add 16 µL of each standard (Std 7 to Std 0) and 16 µL of sample into their dedicated wells (according to the plate map).

8

Dispense the mixed antibody solution.
Add 4 µL of the mixed antibody solution to all wells.

9



Seal the plate and incubate at room temperature.
*Following incubation, the signal remains stable over a period of 24 hours.

10

Remove the plate sealer and read on an HTRF compatible reader.

See assay recommendations on the back

SHORT PROTOCOL - RECOMMENDATIONS

- We recommend running the standard curve in triplicate wells.
- Store the kit at ≤-60°C. Under proper storage conditions, reagents are stable until the expiry date indicated on the label.
- Once reconstituted, standard stock solution may be frozen, and can be thawed only once.
- Once thawed, antibody solutions can be frozen once.
- To avoid freeze/thaw cycles, it is recommended to dispense remaining stock solutions into disposable plastic vials for storage at ≤-60°C.
- Volume of IL8 standard and antibody aliquots should not be under 10 µL.
- Thawed diluent and detection buffer can be stored at 2-8°C in your premises. It is very important to prepare reagents in the specified buffers. The use of an incorrect diluent may affect reagent stability and assay results.
- Thaw the frozen reagents at room temperature.
- Before use, allow antibodies, diluent and detection buffer to warm up at room temperature then
  - homogeneize buffer and diluent with a vortex
  - centrifuge (NEVER vortex) the antibodies to gather all liquid at the bottom of the vial
- It is recommended to filter buffers before use.
- Prepare the standard curve in the diluent provided in the kit or in the cell culture medium used to prepare your samples. In order to prevent any sticking, the cell culture medium must be supplemented with BSA, 10 % FCS or any other carrier protein.
- Due to the stability of the IL8, it is mandatory to prepare the standard curve just before the assay.
- In order to check for a potential interference effect from your assay buffer when using the assay for the first time, we highly recommend the parallel preparation of a standard curve in your own supplemented cell culture medium and in diluent.
- In order to counteract any standard sticking we recommend changing tips between each dilution.

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