

ASSAY PROTOCOL

- Dilute the anti GST-Donor Ab 50-fold with Binding Domain Detection Buffer #1 to obtain the working solution ready to be dispensed.
- The peptide-biotin / streptavidin-acceptor ratio must be equal to 8/1 final in the well (e.g. Peptide-biotin used at 4 nM final in the well, SA-Acceptor must be used at 0.5 nM final in the well). Prepare the SA-Acceptor solution in Binding Domain Detection Buffer #1 to get a 4X working solution depending on the final optimal concentration in the well.

Prepare working solutions of protein and biotin-peptide in assay buffer just prior to use.

- We recommend using the GST-tagged binding domain at 5nM final concentration in the well. Prepare the working solution at 5X depending on the final concentration in the well in Binding Domain diluent buffer (here 25 nM).
- Prepare the peptide-biotin at optimal concentration (referenced in table Optimal experimental conditions) in Binding Domain Diluent Buffer to get a 5X working solution depending on the final optimal concentration in the well.
- Prepare supplemented Binding Domain Diluent Buffer with DMSO to get a constant percentage throughout the inhibitor titration. Dilute the compound in this solution to get a 10X working solution depending on final concentration in the well.

DMSO may act as an inhibitor of GST-binding domain and the biotinylated peptide interaction. This can lead to a decrease of the assay window as DMSO % increases. We recommend the use of compatible DMSO % (See table "Optimal experimental conditions" for DMSO tolerance associated with CBX1).

- Add to a 384-well small volume plate in the following order:
 - 4 µL of CBX1, GST-tag (5X)
 - 2 µL of assay buffer (w/ or w/o DMSO)
 - 4 µL of [Lys(9)Me3] H3(1-21)-biotin (5X)
 - 5 µL of SA-Acceptor (4X)
 - 5 µL of anti GST-Donor Ab (4X)
- Cover the plate with a plate sealer and incubate 1h at room temperature.
- *Remark: Signal remains stable after Over Night incubation.*
- Remove plate sealer and read fluorescence emission at 665nm and 620nm wavelengths on an HTRF compatible reader *.

	PEPTIDE TITRATION		TEST OF INHIBITORS		
	POSITIVE SIGNAL	NEGATIVE CONTROL	INHIBITOR	POSITIVE CONTROL	NEGATIVE CONTROL
CBX1, GST-TAG	4 µL	-	4 µL	4 µL	-
INHIBITOR	-	-	2 µL	-	-
BINDING DOMAIN DILUENT BUFFER	2 µL	6 µL	-	2 µL	6 µL
BIOTIN-PEPTIDE	4 µL	4 µL	2 µL	4 µL	4 µL
STREPTAVIDIN-ACCEPTOR	5 µL	5 µL	5 µL	5 µL	5 µL
ANTI GST-DONOR AB	5 µL	5 µL	5 µL	5 µL	5 µL

* For more information on HTRF compatible reader, please visit www.cisbio.com/htrf-compatible-readers.

OPTIMAL EXPERIMENTAL CONDITIONS

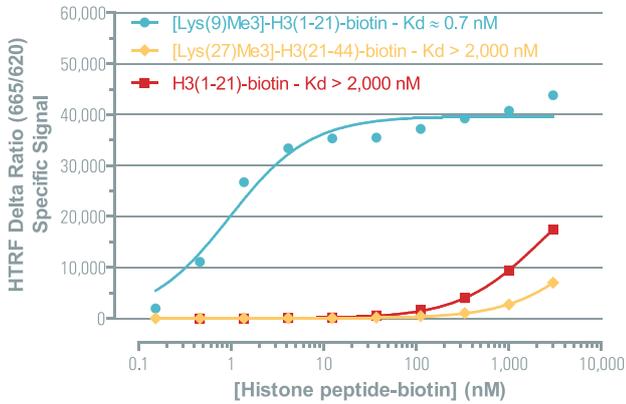
BINDING DOMAIN	RECOMMENDED PEPTIDE CONCENTRATION (FINAL IN THE WELL)	DMSO TOLERANCE
CBX1	0 - 4% DMSO: 1 nM	0 - 4%

DATA REDUCTION

- The TR-FRET signal is treated as HTRF Ratio = Acceptor signal (665nm) / Donor Signal (620nm) x 10⁴
- HTRF Delta Ratio = Ratio (Positive) – Ratio (Negative) where Negative control is performed without reader-protein.
- Assay window = S/B = Ratio (Positive) / Ratio (Negative)

RESULTS

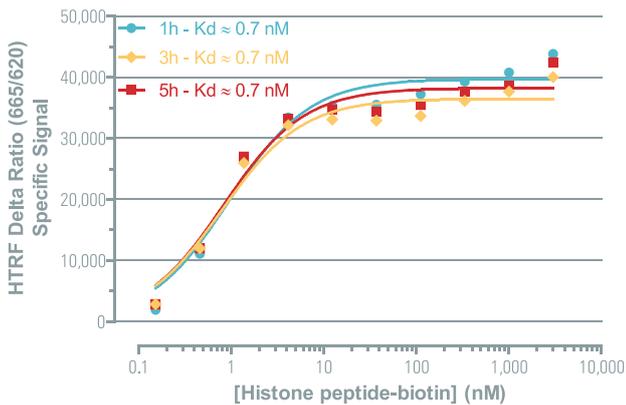
1. PEPTIDE-BIOTIN TITRATION



Measurement of CBX1 / histone H3 peptide interaction and peptide selection

The GST-CBX1 concentration was fixed at 5 nM while the peptide-biotin was serially diluted. The HTRF Delta Ratio is proportional to the specific interaction measured between GST-CBX1 and peptide-biotin. The 0.7nM K_d value for [Lys(9)Me3]-H3(1-21)-biotin peptide is determined from this experiment using a one-site specific binding regression. As expected, unmodified histone peptide-biotin and K27Me3 methylated peptide display very low specific binding compared to K9Me3 peptide (Jacobs et al. EMBO, 2001 and Kaustov et al. Jbc, 2011).

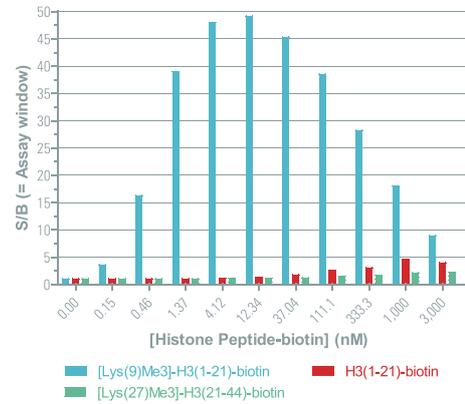
3. KINETIC OF PEPTIDE-BIOTIN BINDING



Measurement of CBX1 / histone H3 peptide interaction kinetic

The GST-CBX1 concentration was fixed at 5 nM while the peptide-biotin was serially diluted. The HTRF Delta Ratio is proportional to the specific interaction measured between GST-CBX1 and [Lys(9)Me3] H3(1-21)-biotin peptide. Equilibrium of kinetic binding of peptide-biotin on bromodomain is reached at 1h.

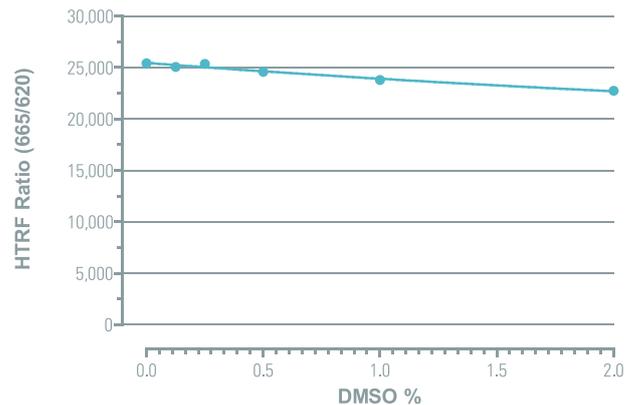
2. ASSAY WINDOW WITH VARIOUS PEPTIDE-BIOTIN



Selection of optimal peptide-biotin and concentration

The [Lys(9)Me3]-H3(1-21)-biotin peptide which gives the best results was selected at 1nM for inhibitor titration. Note that the higher the peptide-biotin concentration, the higher the inhibitor IC50.

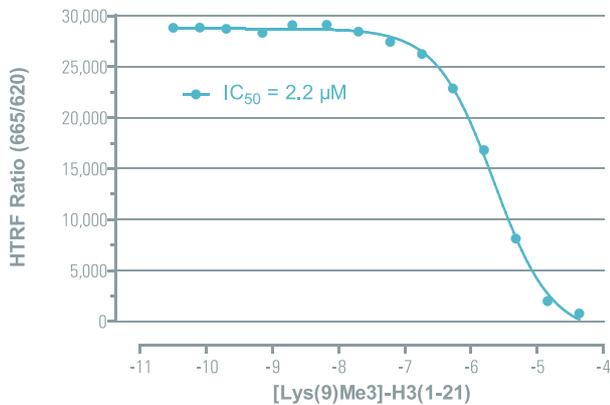
4. DMSO EFFECT



CBX1 HTRF assay displays very good DMSO tolerance

The HTRF assay was performed using 2 nM peptide-biotin, 5 nM GST-CBX1 and DMSO ranging from 0 to 2%. No major DMSO effect is observed on signal.

5. INHIBITOR TITRATION



CBX1 HTRF inhibition assay was validated using reference inhibitors

The HTRF assay was performed using 1 nM peptide-biotin, 5 nM GST-CBX1 and 1% DMSO set constant throughout the inhibitor titration. The IC₅₀ of [Lys(9)Me₃]-H₃(1-21) peptide is in good agreement with published data (Kaustov et al. Jbc, 2011 - 5 μM)

For more information, please visit us at www.cisbio.com/epigenetic-binding-domain

RELATED INFORMATION

Enabling epigenetics studies from HTS to SAR: a novel HTRF® platform to identify and characterize reader domain inhibitors

Roux T, Badol M, Douayry N, Sergeant L, Trinquet E, Degorce F, Milhas S, Betzi S, Derviaux C, Eydoux C, Letienne J, Lugari A, Collette Y, Guillemot J-C, Morelli X. -

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How do HTRF® epigenetic binding domain assays perform compared to other technologies?

Thomas Roux, Najim Douayry, Laurent Sergeant, François Degorce and Eric Trinquet.

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